#4169 CReMEC initiative: Characterization of patient-derived colorectal tumor models and correlation with patient profile



CREMEC

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p = 0.0292 (*)

n = 0.0287 (*)

Well (n=9)

INTRODUCTION

Well characterized models representing the heterogeneity of human colorectal cancers (CRC) are needed to develop effective therapeutic agents. Establishment of such tools will allow a better prediction of the clinical outcome, taking into account the diversity of each patient tumor phenotype and genotype. For this purpose, we have associated efforts from hospitals, academic groups, biotech and pharmaceutical companies. The goal of this consortium is to create an experimental tumor model resource center to improve or strengthen drug development. From May 2007 to January 2009, 86 surgical specimens [59 primary (P) tumors, 19 metastasis (M), and 8 peritoneal carcinomatosis (C)] were collected from CRC patients (with informed consents and negative HBV, HCV, and HIVs serologies). Tumor samples were subcutaneously xenografted in *nude* mice and characterized as described below. We report here the results on our panel of models.

MATERIAL AND METHODS



Patients were informed and gave their consent for providing surgical tumor samples to CReMEC and for HIV1, HIV2, HBV and HCV serological status testing. Tumor samples were collected in 3 medical centers of the Paris-Ile-de-France area: Institut Curie, Institut Gustave Roussy, Hôpital Lariboisière. Fresh tumor material was conditioned into RPMI 1640 with 200 U/mL penicillin, 200 µg/mL streptomycin and 2.5 µg/mL nografting was carried out at the site of sampling within 12 hours after specimen collection. Procedure were performed according to ethical guidelines for anima

care and handling.

20-40 mg fragments were xenografted subcutaneously either in the flank or in the interscapular area, in 3-6 immunodeficient SWISS *nude* mice Grafted mice were kept for a maximum of 4 months withou

Clinical data collection

Relevant clinical information was collected by the attending physician and included in a standardized data sheet Identification of the data sheet is anonymous

Histological characterization

Samples were fixed for a maximum of 48 hours in alcohol-formalin-acetic acid (AFA) and embedded in paraffin. 5 um section

Molecular characterization - sequencing

- CGH array analysis: Evaluation of genome-wide, gene copy number was evaluated using a 244k CGH array Agilent technology. It was carried out on DNA from the patient sample or at an early passages (P0/P1), and at the passage used for the pharmacological evaluation (P8-9).
- DNA sequencing: The following genetic markers, relevant in CRC, were selected for sequencing: APC (exons 9 & 16), KRAS (exons 2 & 3), BRAF (exons 11 & 15), TP53 (exons 2 to 11), CTNNB1 (exon 3), PIK3CA (exons 10 and 21), FBXW7 (exons 4 to 11), EGFR (exons 18 to 21) and AKT1 (exon 4). Sanger direct sequencing was performed after PCR amplification of exons of genes harboring hotspot mutations as described in Catalogue Of Somatic Mutations In Cancer (COSMIC: DNA se /cosmic/).
- Determination of Microsatellite Instability (MSI) status: The MSI status was determined according to the Consensus Conference (or Revised Bethesda Guidelines) (J. Natl Cancer Inst 2004, 96, 261-268) recommendations using the following five quasimonomorphic markers: NR21, BAT26, BAT25, NR24 and NR22.

Pharmacological characterization

- Tested agents: 5-fluorouracil (5-FU), oxaliplatin (L-OHP), irinotecan (CPT-11) or cetuximab, were tested.
- In vivo determination of antitumor activity: Tumor fragments were subottaneously xenografted in SCID mice or nude rats. Tumor-bearing animals were randomized when the mean tumor volume reached 100-200 mm³ in mice or 500-700 mm³ in rats. Compounds were formulated in glucose 5% in water. 3 cytotoxic drugs were IV administered, and tested at 70% of their respective highest non toxic dose and using the following regimen in mice: 5-FU at 56 mg/kg/adm QTDx2 (30 mg/kg/adm QTDx3 in rats), L-OHP at 5 mg/kg/adm Q4Dx2 (4 mg/kg/adm Q4Dx3 in rats), CPT-11 at 22 mg/kg/adm, Q2Dx3 (40 mg/kg/adm, Q7Dx3in) rats) ; 1 targeting therapy was IP administrated: cetuximab at 12.5 mg/kg/adm, (Q3Dx2)x2 (10 mg/kg/adm, IV, Q7Dx3 in rats).

Antitumor activity was evaluated by calculating the $\Delta(1/C)$ ratio:			
Scoring criteria:			
	median T vol. py - median T vol. py	- = Δ(T/C) > 42 %	
∆(T/C) (%) = -	VOL-D1 VOL-DX X 100	+ = 10 < Δ(T/C) ≦ 42 %	PR: Partial Response
	median C VOL-DY - median C VOL-DX	++ = 0 $\leq \Delta(T/C) \leq 10$ % (stable disease)	CR: Complete Response
		$+++ - \Lambda(T/C) < 0 \%$ (tumor regression)	TES: Tumor Free Survival



Significant distribution^{*} of primary tumors in regards of lymph node status (panel A, N1 = 1 to 6, N2 = 7 to 15 positive regional lymph nodes), stages (panel B) and differentiation status (panel C)

No other significant correlation were found among the following parameters: gender, age, resection extent, lymphatic embolies, perinervous invasion, initial treatment, genotype * p values were calculated with logical regression te





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----- Plan to complete the full correlation analysis between clinical data, gene mutations, transcriptome profile, ex-vivo and in vivo drug sensitivity. Perspectives to fully exploit this new collection for new drug candidate selection.