Biological characterization and non invasive PET imaging explorations of lymphatic dissemination in a human melanoma xenograft model

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Olivier DUCHAMP¹, Raphaël BOISGARD², Bertrand TAVITIAN², Philippe GENNE¹ and Nicolas GUILBAUD¹



1) Oncodesign SA, Dijon, France; 2) In vivo imaging of gene expression laboratory, INSERM/CEA/SHFJ, Orsay, France

INTRODUCTION

The major cause of death from melanoma is related to metastases development, these secondary tumours being resistant to conventional therapies. Determining the metastatic spread of melanoma cells to the regional lymph nodes is essential in the assessment of prognosis and treatment options. Several human melanoma cell lines have been reported to metastasize in the immune-deficient mice, but lymph nodes metastatic rates were extremely low. To evaluate the potential of therapies against metastatic melanoma, we developed a novel human xenograft model in immunodeficient rodents (mice and rats) involving metastasis formation in multiple organs

Positron emission tomography (PET) is a diagnostic imaging modality that allows the characterisation of disease based on altered metabolism. ¹⁸F-fluorodeoxyglucose (FDG) is the most frequently used PET radiotracer. The sensitivity of the microPET® scanner led to the suggestion that FDG-PET might be used i) for initial non-invasive spreading assessment and, ii) for pharmacological treatment investigation in our experimental melanoma model.

OBJECTIVES

In vitro and in vivo biological characterization of the CMEL-5 human melanoma cell line

To investigate the sensitivity of the CMEL-5 xenografts models to reference drugs To evaluate the usefulness of PET imaging modality for the melanoma spreading and for the tumour response

after BCNU treatment on nude rats

ORIGIN OF THE CMEL-5 MELANOMA CELL LINE

The LB1319-MEL cell line was purchased from Dr. B. Van Den Heynd (Ludwig Institute, Brussels) and was first established in vitro from a metastatic malignant melanoma in a 72-year old caucasian male. The CMEL-5 human melanoma cell line was originated from a brain metastasis induced by LB1319-MEL cells intravenously (IV) injected in nude mice. The CMEL-5 cell line was obtained by in vivo selection for its high

capacity to lymph nodes tropism when injected IV to nude mice. METHODOLOGY

Immunocytometry analysis of the CMEL-5 cell line

CMEL-5 cells were grown as monolayer in complete culture medium (RPMI1640/10% FBS). Cells were trypsinized for cell suspension obtaining and 10⁶ cells were incubated with a panel of antibodies to evaluate by Flow cytometry (FACScan) the expression of AlphaVBeta3 (LM609, Upstate, Charlottesville, USA), VEGFR2, IL-18 Receptor, VLA-4, CCR7 (R&D Systems, Mineapolis, USA), and CXCR4 (R&D Systems, Mineapolis, USA).

Experimental tumour models:

Female swiss-nu/nu mice and NIH-rnu/rnu rats 5-7 weeks old of age (Charles River, France) were used for these studies. To develop the disseminated tumour model 106 or 5.106 CMEL-5 cells were intradermaly (OT) injected in Nude mice and Nude rats respectively. To develop the orthotopic tumour model 5.10⁶ or 10⁷ CMEL-5 cells were IV injected in Nude mice and Nude rats respectively. The day of cancer cells injection was considered as the DO

In vivo tumour growth inhibition studies

The tumour sensitivity to chemotherapeutic agents such as carmustin (BCNU), temozolomide (TMZ), 5-Fluorouracil (5-FU) and taxol (TXL) was investigated. Nude mice and Nude rats bearing CMEL-5 orthotopic (OT) or disseminated (IV) tumours were randomized at D13-D21 according to tumour volume (for OT models) or to body weight (for IV models) to constitute groups of 5-6 animals. The treatment doses and schedules were indicated in the table below:

Animals	Tumour implant. site	Treatment	Treatment doses	Treatment adm.	Treatment schedule	Treatment start after cells graft
Nude mice	OT	BCNU TMZ 5-FU TXL	20 mg/kg/inj 50 mg/kg/inj 80 mg/kg/inj 15 mg/kg/inj	IV PO IV IV	Q7Dx3 Q3Dx5 Q7Dx3 Q7Dx3	D13 D19 D19 D19 D19
Nude rats	OT	BCNU	20 mg/kg/inj	IV	Q7Dx3	D19
Nude rats	IV	BCNU	10 mg/kg/inj	IV	Q14Dx2	D21

Positron emission tomography

The PET imaging studies were performed on Nude rats bearing IV induced CMEL-5 tumours on a microPET® FOCUS 220 scanner (Concorde Microsystems-CTI) which has a spatial resolution of 1.34 mm at the center of the field of view. Rats were injected with 37 MBq of ¹⁸FDG via the tail vain. 45 min after injection, the rats were placed in the centre of the camera's field of view, and PET images were acquired during 15 min by bed position (3 positions for whole body acquisition). Images were reconstructed using the FORE-OSEM 2D algorithm and analyzed using the Asipro VM analysis tool (Concorde Microsystems- CTI). PET diagnostics were performed from D13 to D117 for 8 Nude rats and from D40 to D97 for the 3 BCNU treated rats





IV-induced CMEL-5 tumour development in the Nude rats

Time after	IP lymph	Ovaries	Brain	Bone	Lung
Cells ini.	nodes	Adrenal gland			
D20-D30:	0 / 0	0 / 0	0 / 0	25 / 12	25 / 0
D30-D50-	80 / 0	0 / 25	0/0	40 / 37	40 / 0
200 200.	,	0,20	0,0	,	,
D50-D120:	100 / 12	80 / 25	60 / 0	60 / 37	20 / 0





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4: BCNU in IV-induced

FDG-PET study of BCNU efficacy in *Nude* rats

Groups	Nb of rats with positive PET lesions (D50-D75)	Table 2: Rats were IV injected with CMEL-5 cells at D0. Until	
CONTROL	4 / 8	diagnosed with tumour lesions	
BCNU	0 / 3	versus 0% of BCNU-treated	

Belhochine T. et al., The Oncologist, 2002, 7:271-278 Mijnhout GS. Et al., Journal of Clinical Pathology, 2002 Swetter SM. Et al., Annals of Surgical Oncology, 2002, 9:646-653



